# A novel algorithm for discovering transcription factor binding sites

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### 1. Background & Objectives

- Since the advent of the microarray, gene expression over the course of biological processes can be seen.
- **❖**Experimental determination of transcription factors is possible, but slow and expensive: computational methods usually search motifs or gene profiles.(1, 2)
- ♦ We aim to develop a new Matlab tool to search for sites (including pairs) that explain co-regulation.
- **♦**Search for *Arabidopsis* leaf senescence motifs (3).

## 2. Transcriptional Modules (TMs)

- **❖Groups** of genes that are co-regulated in a biological process, by a set of transcription factors.
- **❖Determined** by both their promoter sequence information and time series expression profile.

**ATCAGTGACTTTAACACGGAT** 

**CGAACGTTGATTACGTTGCGT** 

GAAGCTTCTCGGCATTATGCA



### 3. Statistical Sampling

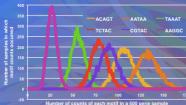


Fig. 1. Bootstrapping returns for six 5-mers in 500 gene proposed TMs.

**♦**5000 samples were taken of the sizes of the TMs generated, to give bootstrap statistics for mean and variance of each motif for each cluster size.

### 4. Elongation & Repetition

- **❖Determine** the most significant motifs via normal approximation (as per Fig. 1.) and extend these in each direction.
- **♦**Use sparsity indicator vectors to reduce the CACCTA size of the search when considering the GACCTA longer motifs. TACCTA



INITIATE: Propose all motifs of a given length k, as initial putative binding sites.

Propose new TMs from the coupled data, using the adapted Kundaje algorithm (4).

Sample from full pool of motif count data sets of proposed transcriptional module sizes. Compare real motif counts for each of the proposed TMs, with bootstrapped values.

Fig. 2. A flow diagram of the steps taken by the algorithm, to determine the putative transcription factor binding site motifs

Calculate the number of each proposed motif present in the promoter of each gene considered.

for motifs via the extension and pairing of the retained motifs. Remove all bar the Nmotifs with greatest specificity for certain proposed TMs.

TERMINATE: Retain putative motifs with highest relevance to the latest proposed TMs.

### 5. Results

- ♦Returned TMs were based upon many micro-satellite repeats.
- ♦Removing pair-repeat 5-mers did not prevent this, but proposed AACCGGTTT as a membrane transport related motif.
- ♦GOstat (5) study of TMs (Fig. 3a, b) revealed more relevance for k-means (Fig. 3c, d) clustering than our algorithm.



**Fig. 3.** Expression profiles of (a,b) Proposed TMs; (c,d) *k*means clusters, for *Arabidopsis* leaf senescence time course expression data (3).





#### 6. References

- 1. Spellman, P.T. et al. (1998) Comprehensive Identification of Cell-Cycle Regulated Genes of the yeast Saccharomyces cerevisiae. Mol. Bio. of
- Tavazoie, S. et al. (1999) Systematic Determination of Genome Network Architecture Nature Genetics 22 281-285. Thanks to the Buchanan-Wollaston Warwick HRI group for their
- Arabidopsis leaf senescence gene expression data set.

  Kundaje, A. et al. (2005) Combining Sequence and Time Expression
- Data to Learn Transcriptional Modules. *IEEE/ACM* 2 (3) 194-202. Beissbarth, T. & Speed, T. P. (2004) GOstat: find statistically overrepresented Gene Ontologies within a group of genes. *Bioinformatics* 20 (9) 1464-1465.



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