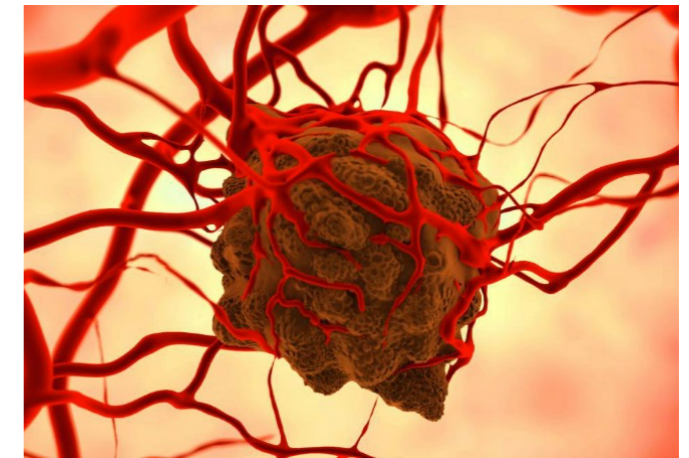


# Leveraging concepts from stochastic simulation and machine learning for efficient Bayesian inference

Ruth Baker

 @ruth\_baker

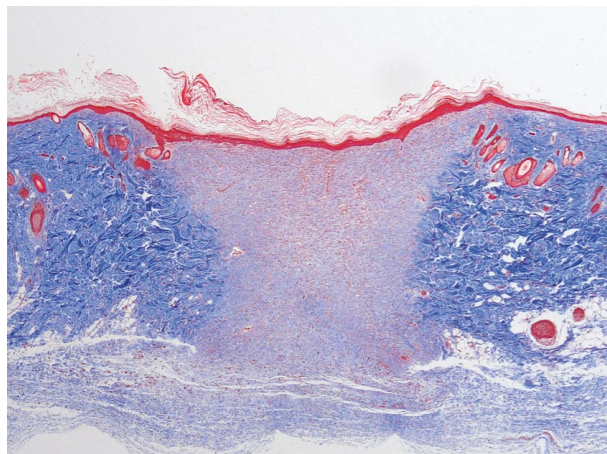
To understand the mechanisms driving collective cell motility, proliferation and death and their contributions to complex biological processes, such as those associated with development, disease and repair.



tumour growth



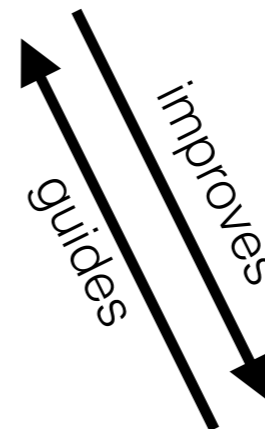
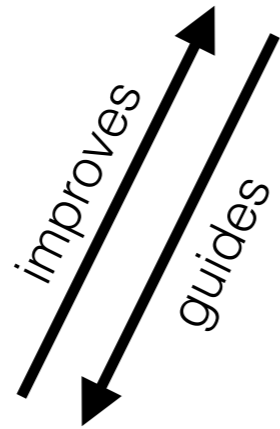
embryo development



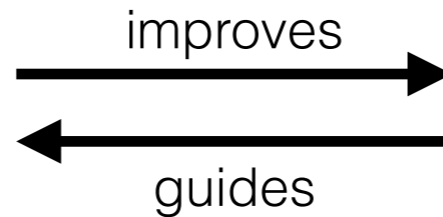
wound healing

Combine mechanistic mathematical models and statistical / machine learning tools to provide new biological insights.

Mathematical modelling  
computational simulations



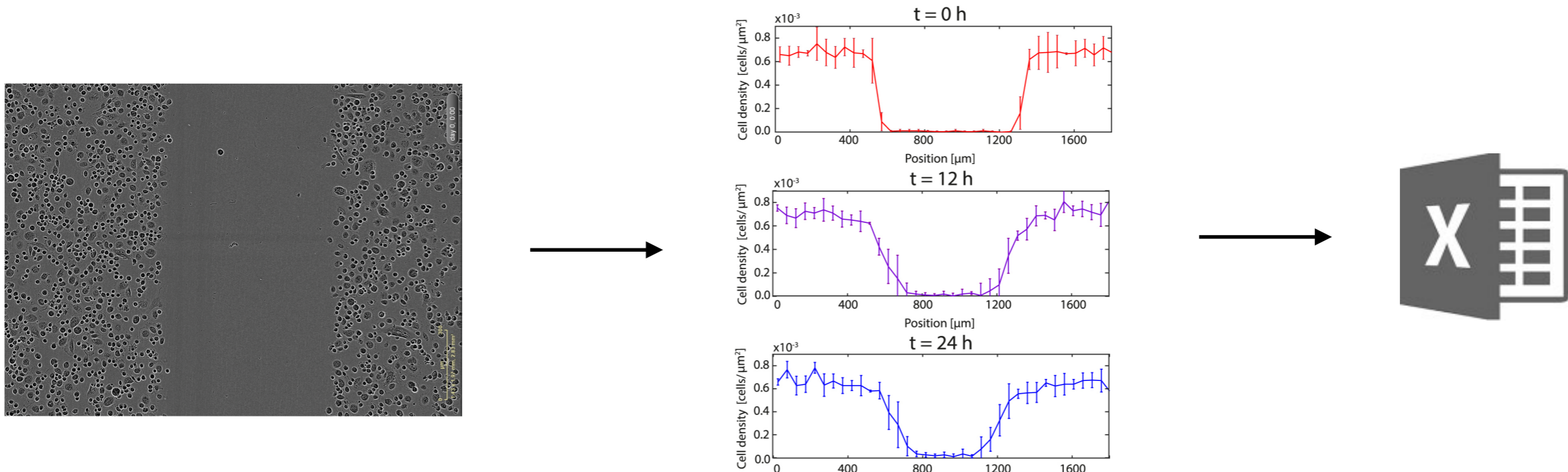
Data analysis and  
model testing



Experiments: wildtype  
and perturbation

Integral in the cycle of  
**predict - test - refine - predict**

- Given quantitative data, can we estimate model parameters?



- Parameter inference using a Bayesian framework:

$$\mathbb{P}(\theta | \mathcal{D}) \propto \mathcal{L}(\mathcal{D} | \theta) \mathbb{P}(\theta)$$

↑                    ↑                    ↙  
posterior    likelihood    prior

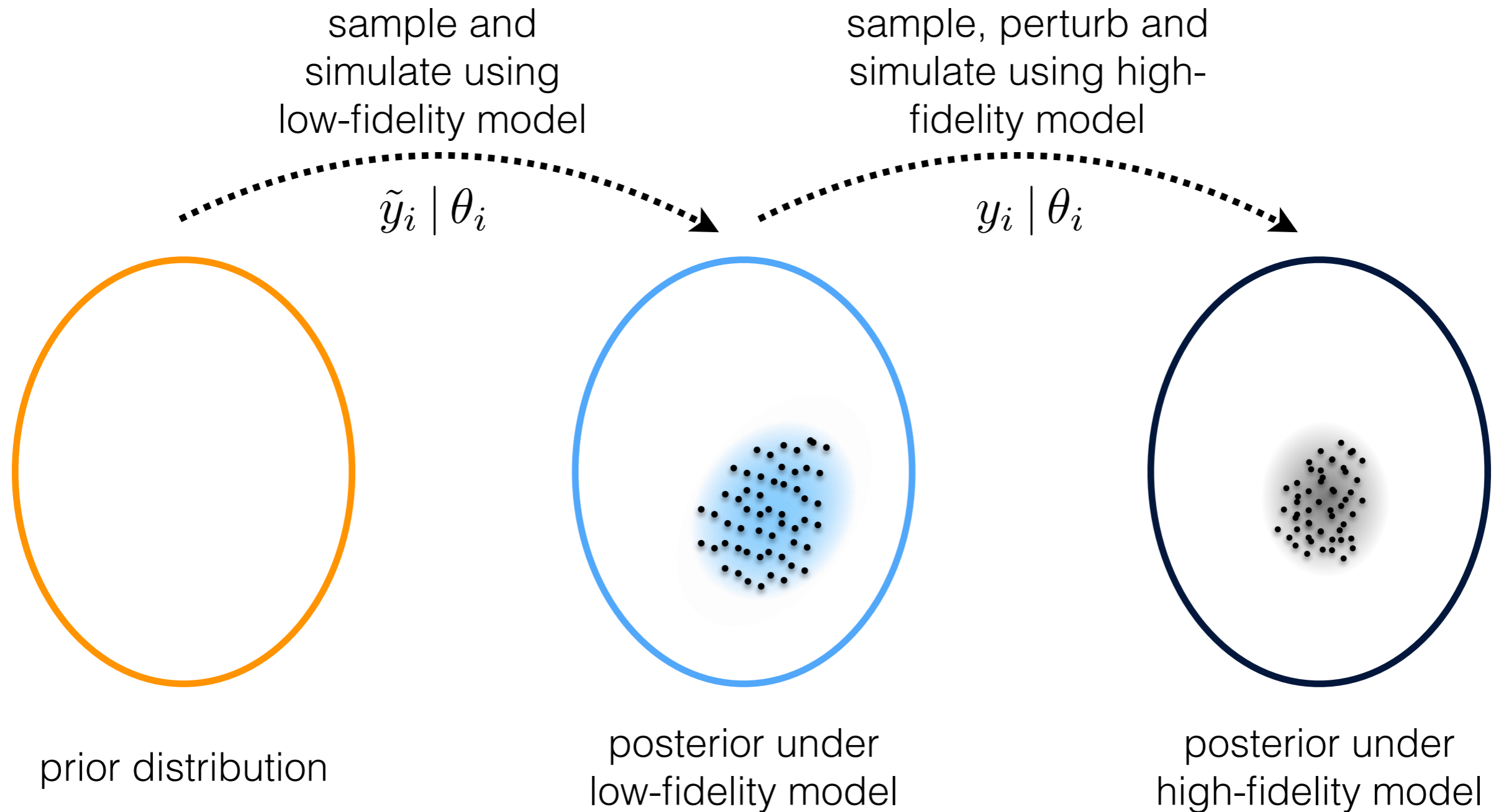
- Likelihood is generally intractable, so we use ABC:
  - A **very large number** of times:
    - sample parameter from some distribution;

**When a model takes minutes (or longer) to simulate, and / or the model has many parameters, ABC methods can be infeasible.**

evaluate how close model output is to data using a summary statistic and distance function;

- assign a weight to the parameter - depends on this distance.

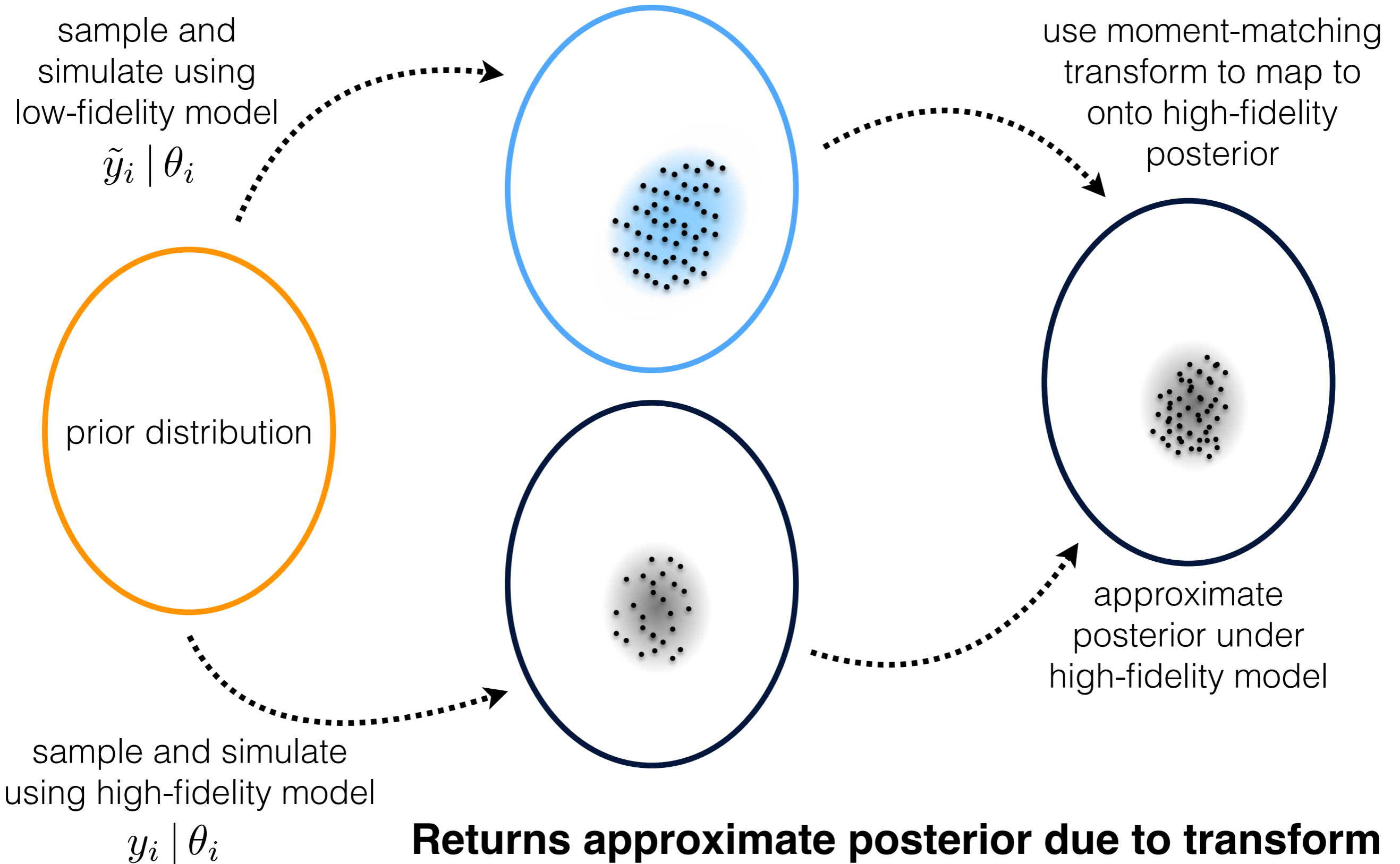
- Use a more intelligent exploration of parameter space.
  - Importance sampling, sequential Monte Carlo *etc.*
  - **Pre-conditioned ABC** - Use a simple model to help transition from prior to posterior.
- Make the weights less expensive to calculate.
  - **Moment-matching ABC** - Use a simple model to help estimate posterior.
  - **Multifidelity ABC** - Use hierarchies of models in the weight calculations.
  - **Minibatch ABC** - Use subsets of the data in evaluating the distance function.



**Weight accepted particles so that ABC posterior is returned.**

Warne, Baker and Simpson, *J. Comp. Graphical Stat.* (2022).

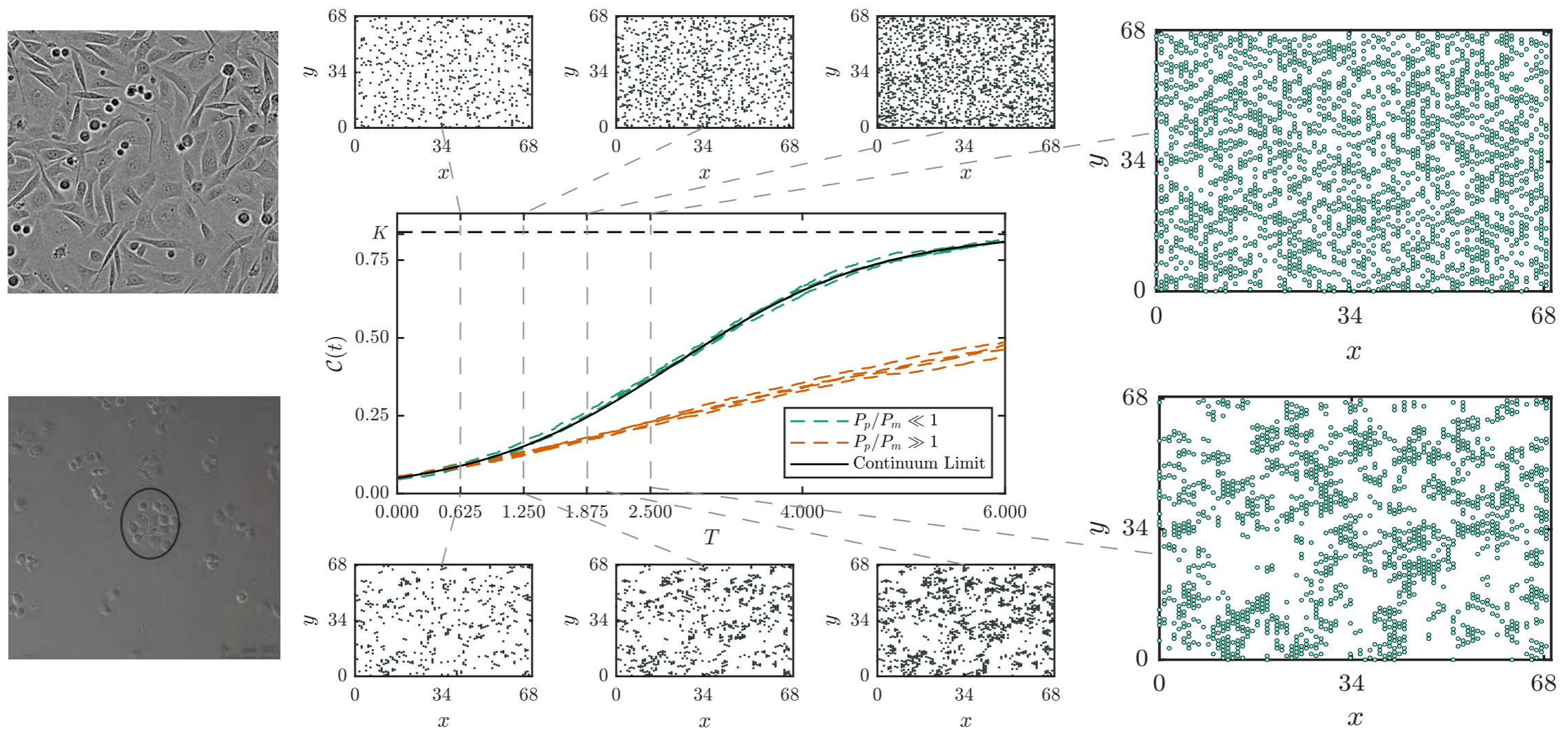
# Moment-matching ABC



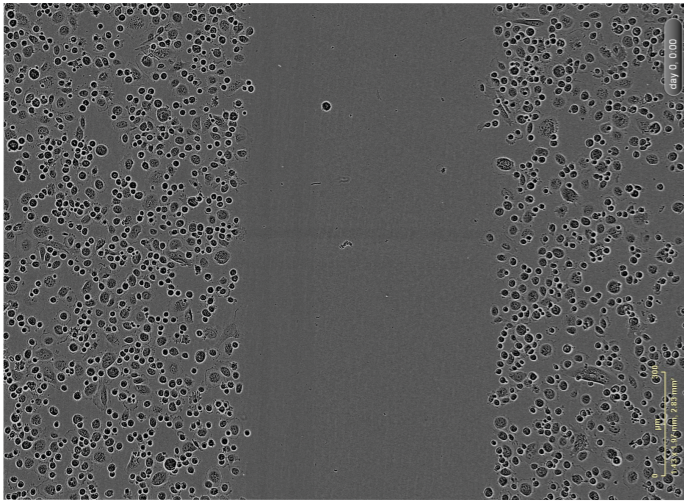


# Pre-conditioned / MM ABC

- Discrete, on-lattice random walk model of cell migration and proliferation.



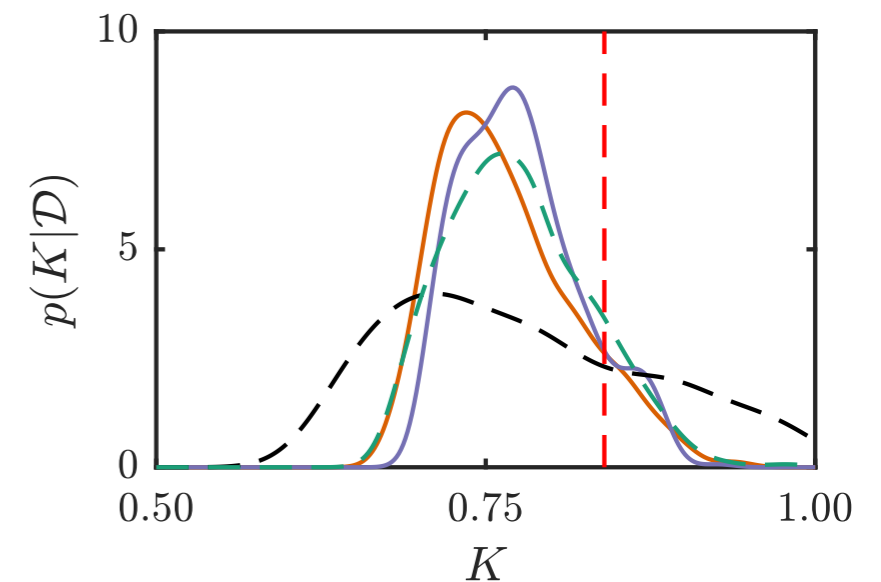
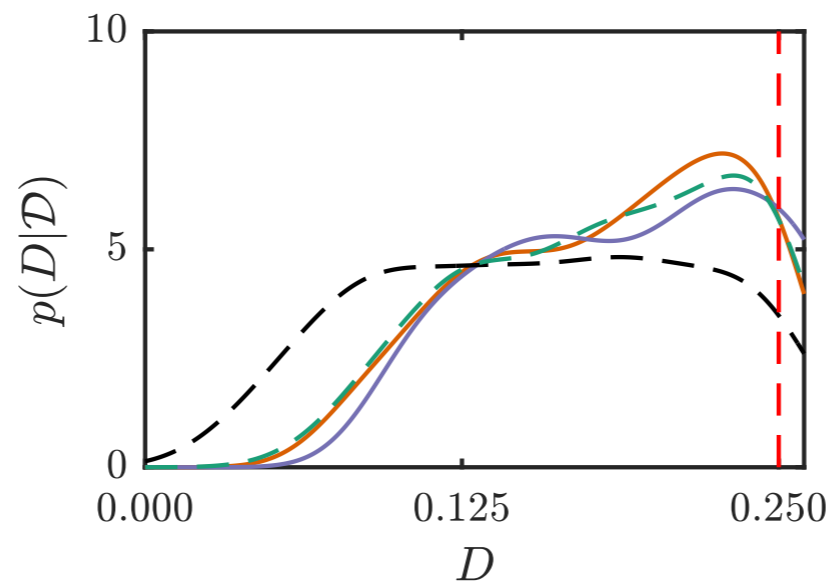
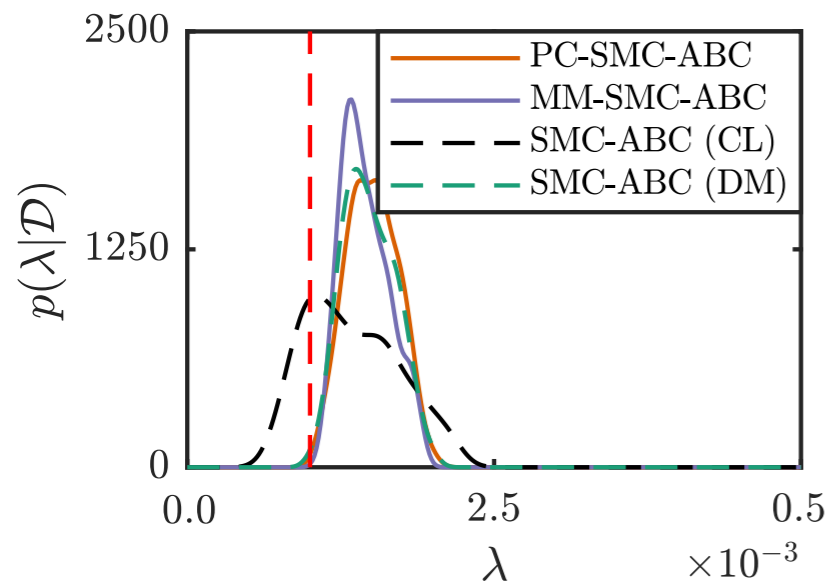
Warne, Baker and Simpson, *J. Comp. Graphical Stat.* (2022).



Continuum limit - Fisher-KPP equation:

$$\frac{\partial C}{\partial t} = D \frac{\partial^2 C}{\partial x^2} + \lambda C \left( 1 - \frac{C}{K} \right)$$

Marginal posterior parameter distributions

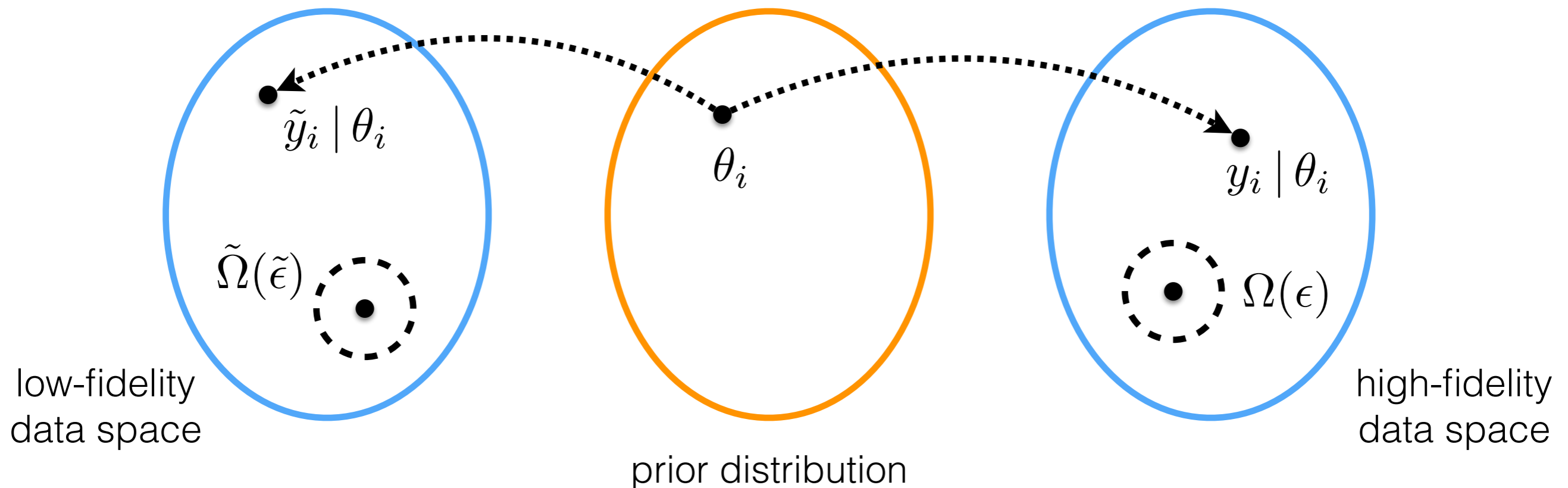


Warne, Baker and Simpson, *J. Comp. Graphical Stat.* (2022).

- Pre-conditioned and moment-matching ABC can provide significant time savings, through the combined use of high-fidelity and low-fidelity models.
- Need to explore trade-off between fraction of high-fidelity and low-fidelity samples.
- Doesn't require the low-fidelity model to be particularly accurate, just that the model outputs depend in a qualitatively similar way on the input parameters.

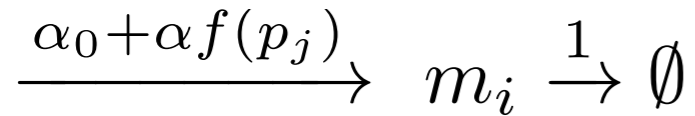
Warne, Baker and Simpson, *J. Comp. Graphical Stat.* (2022).

- Can we use the low-fidelity model to allocate some weights (accept or reject parameters) and not bias the result?

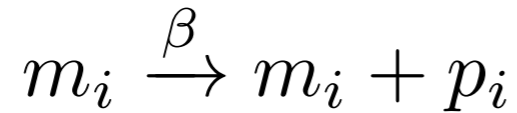


Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).

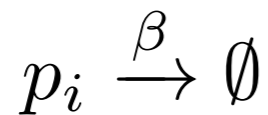
# Multifidelity ABC - example



for  $(i, j) = (1, 3), (2, 1), \text{ and } (3, 2)$

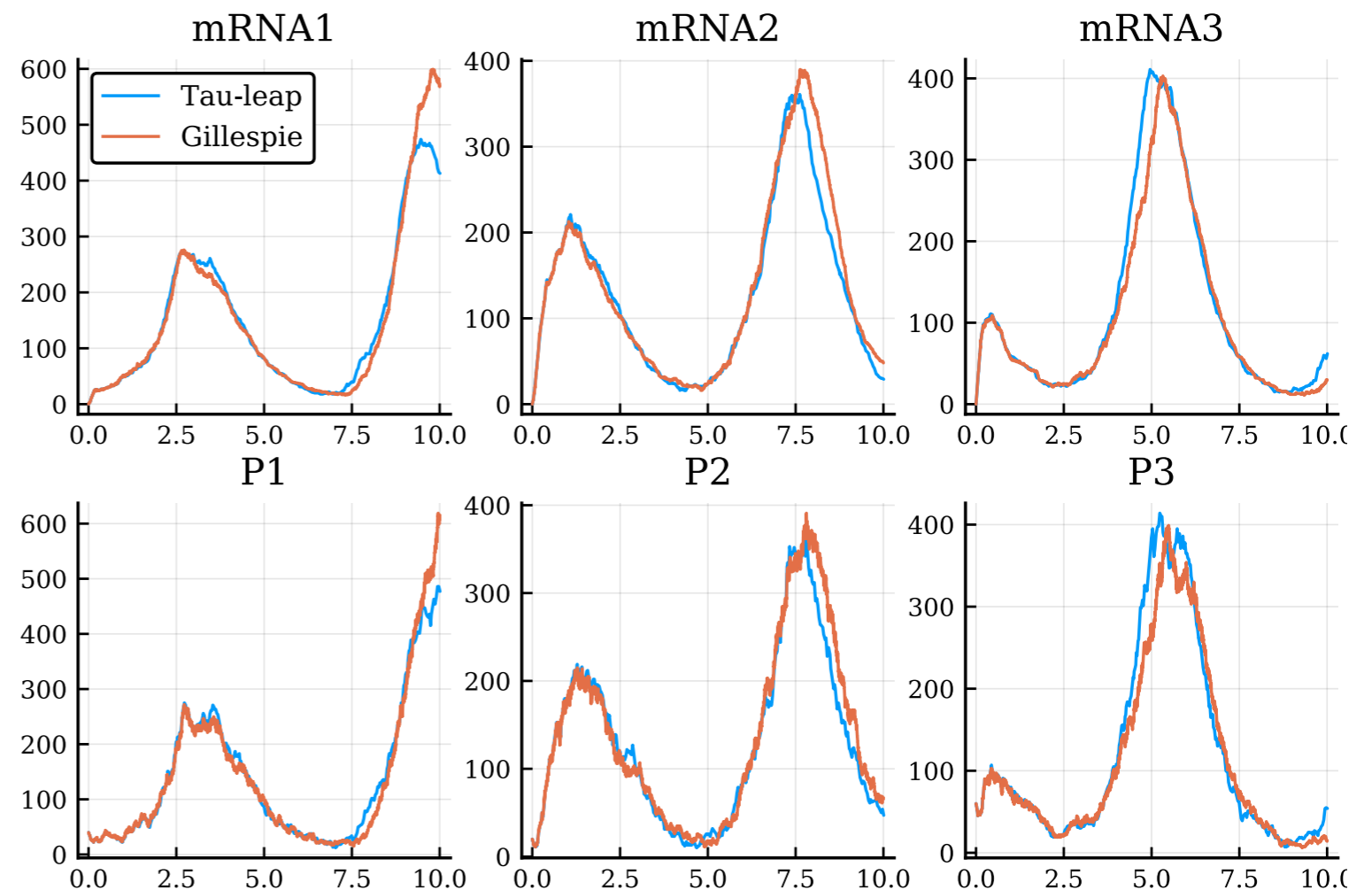
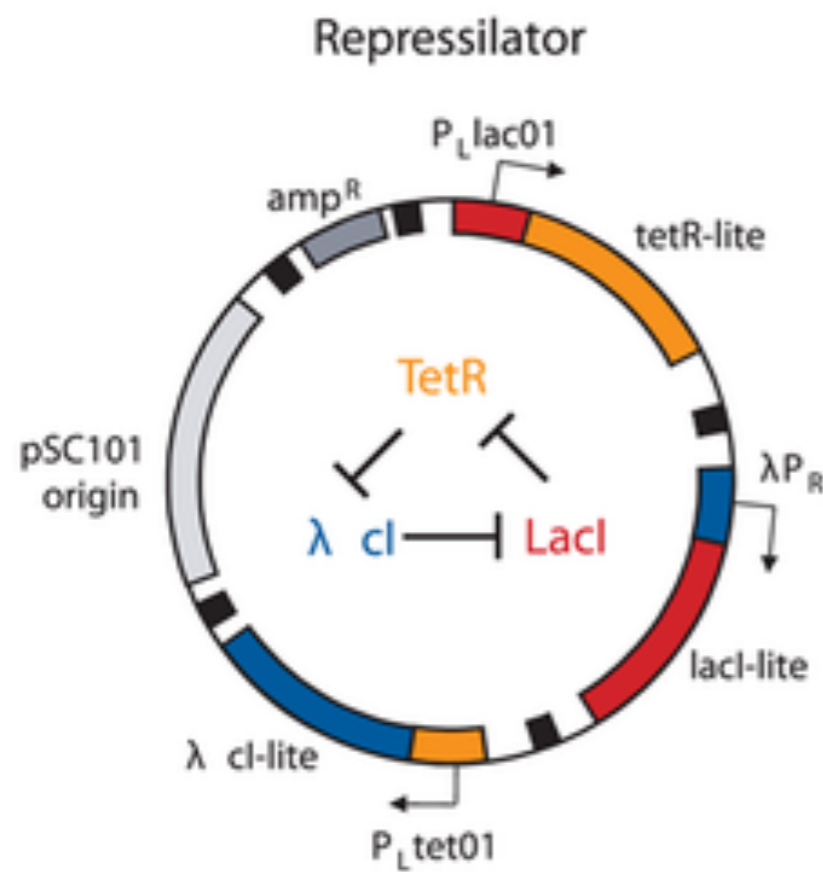


for  $i = 1, 2, 3$

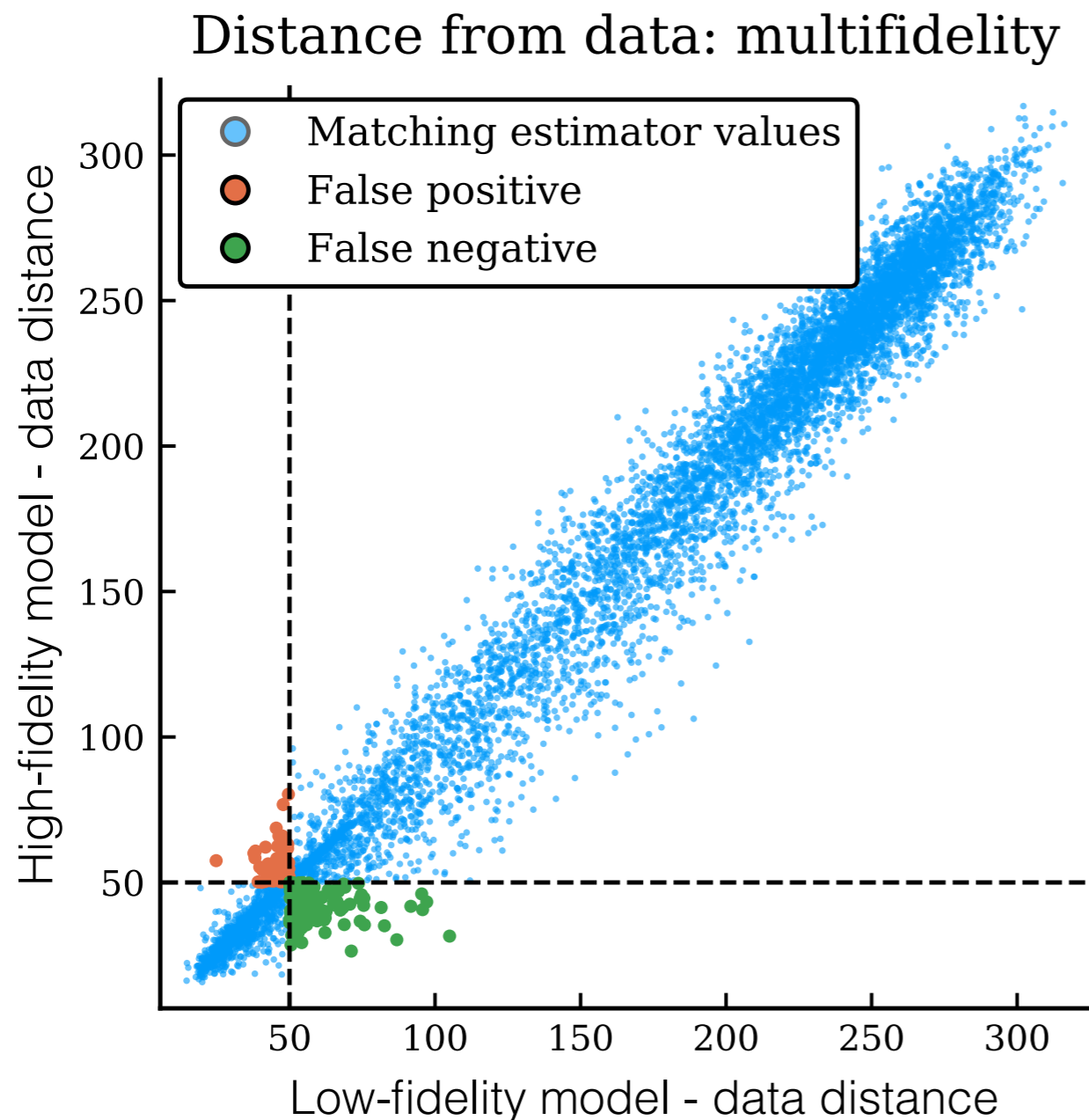


for  $i = 1, 2, 3$

$$f(p) = \frac{K_h^n}{(K_h^n + p^n)}$$



# Multifidelity ABC - the problem



- How to combine the outputs of the two models, so that the result is unbiased weights?
- How can we make this process efficient?

Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).

- Attempt to make an “early decision” using the low-fidelity model, and “sometimes” check that decision using the high-fidelity model.
- Decision to check is made uniformly at random, with probability  $\alpha(\tilde{y}, \theta_n)$ .

- Here, we assume  $\theta_n$  is sampled from a uniform distribution

**Using a common noise input makes this a very efficient approach.**

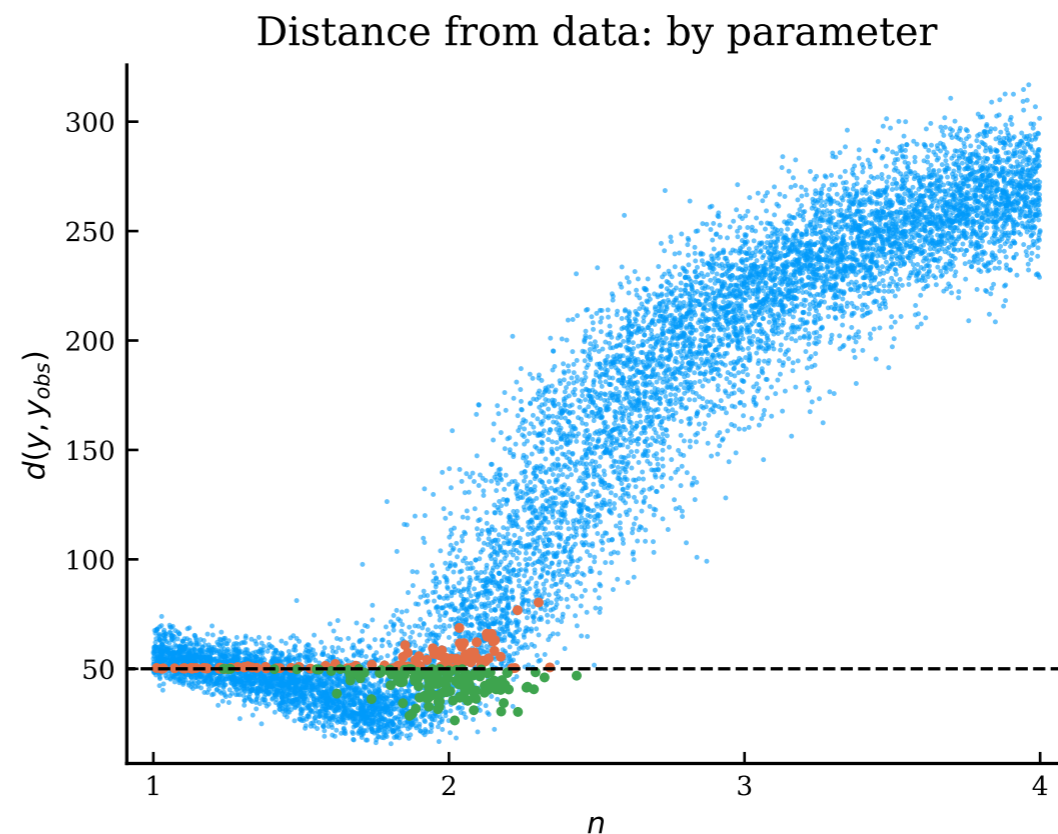
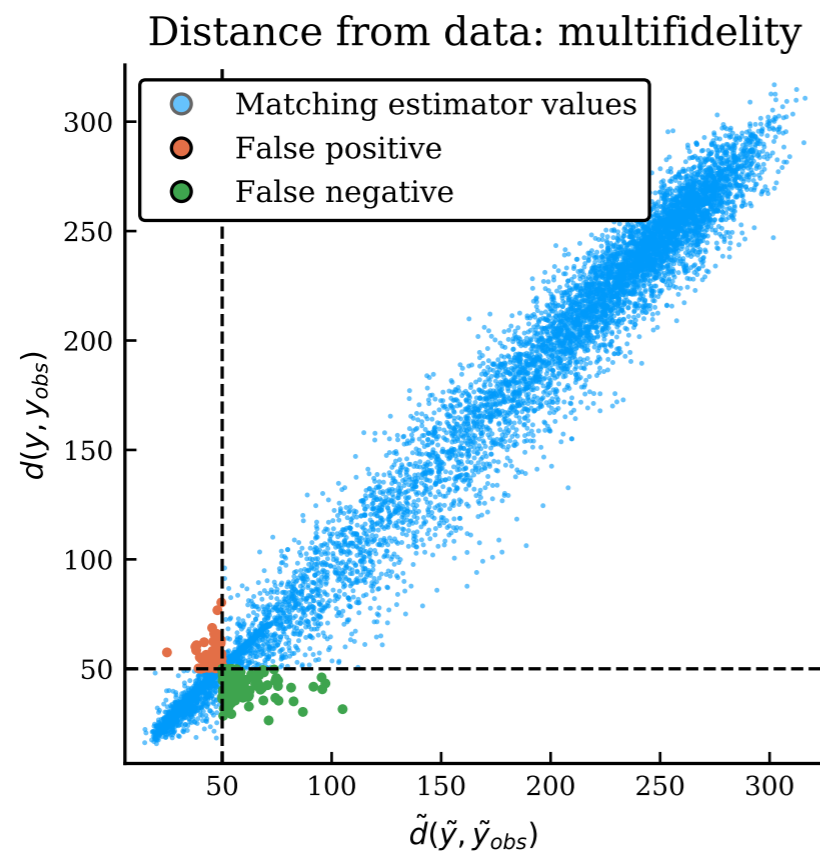
$$\alpha(y, \theta_n) = \eta_1 \mathbb{1}(y \in \mathcal{L}(\epsilon)) + \eta_2 \mathbb{1}(y \notin \mathcal{L}(\epsilon))$$

if low-fidelity model  
is **close to** data

if low-fidelity model  
is **far from** data

Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).

- Derive analytical expressions for the optimal continuation probabilities, given estimates of various quantities.
- In practice: adapt the continuation probabilities “on the fly”, as samples are generated...

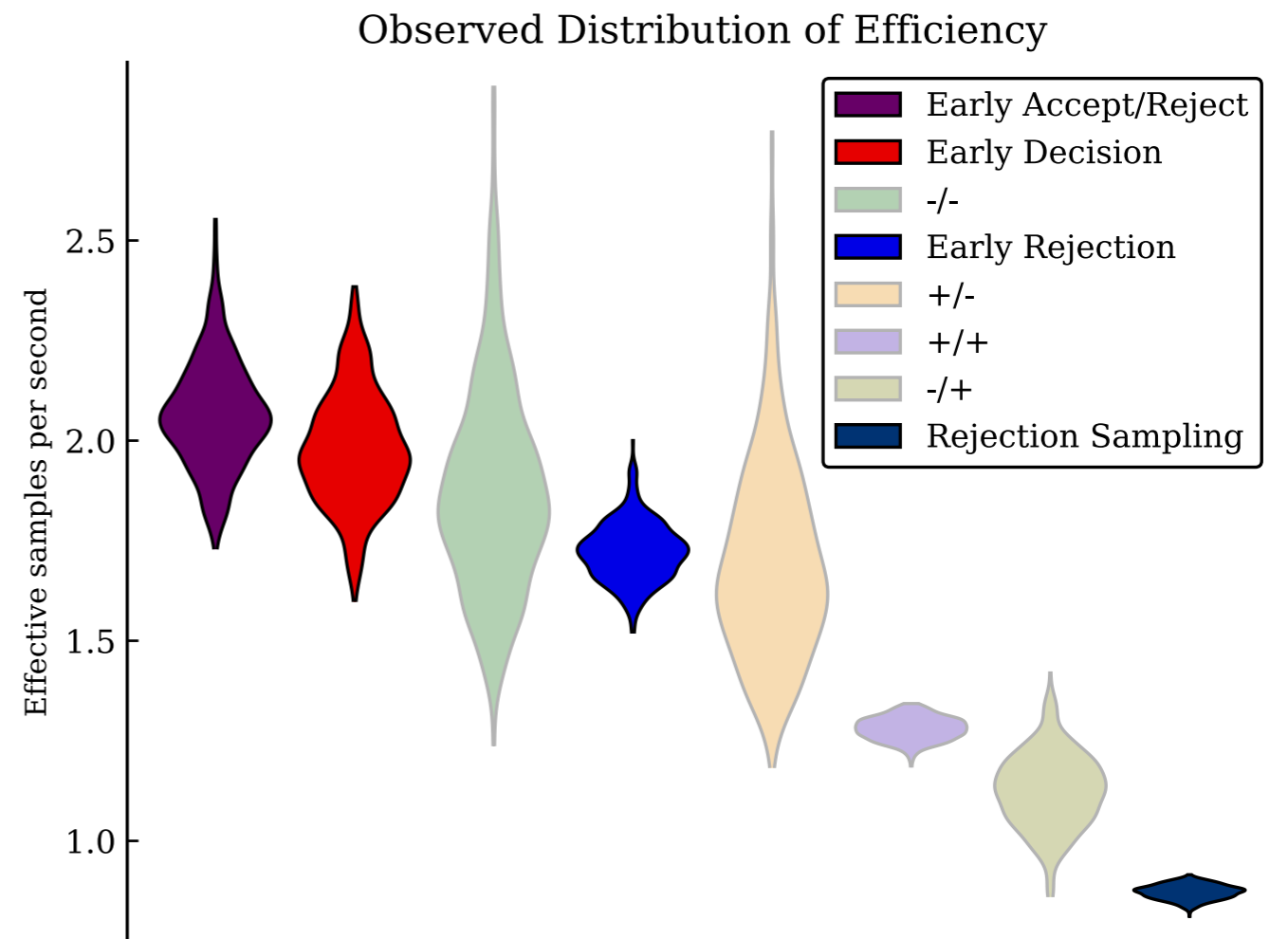
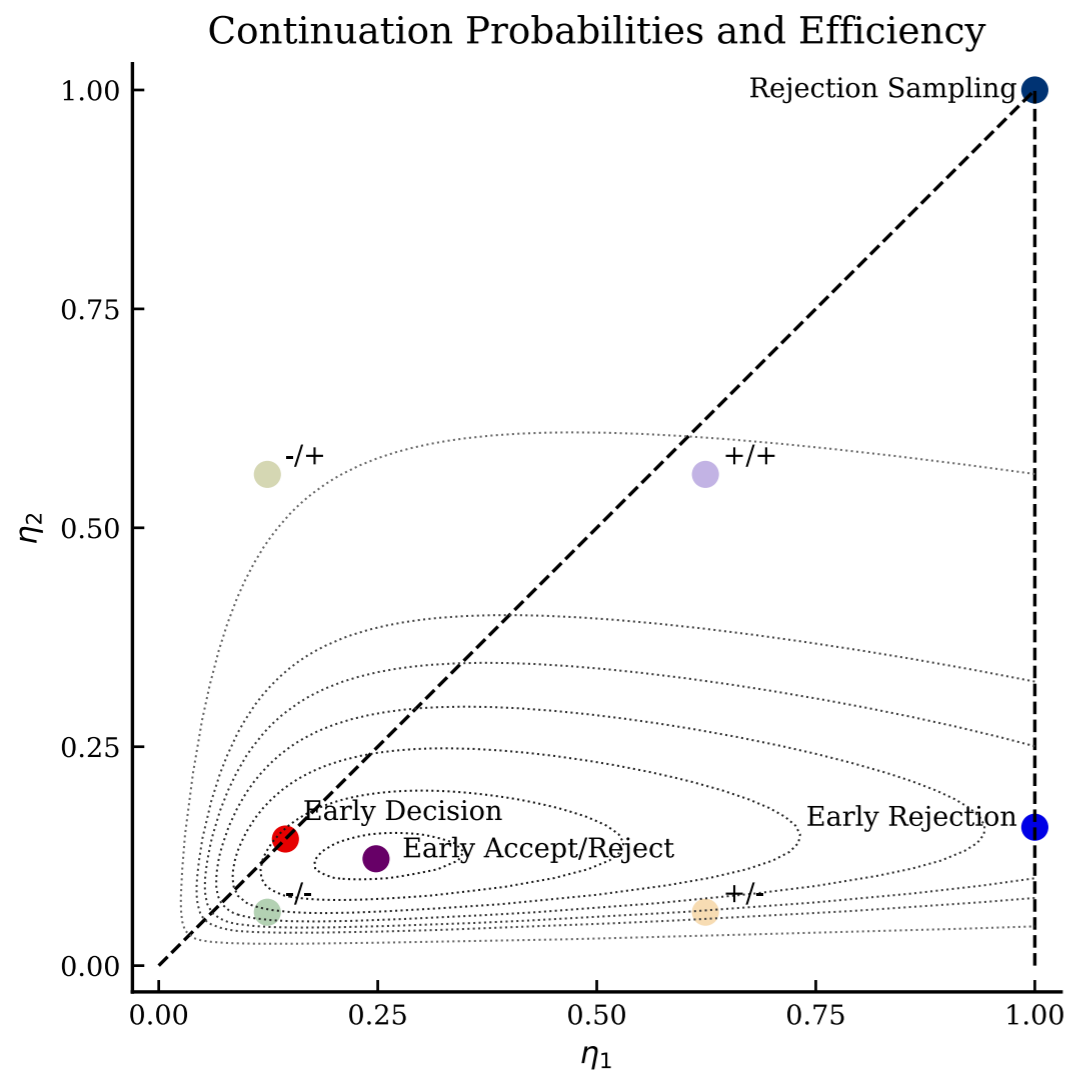


Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).



# Multifidelity ABC - example results

- Comparing results for a range of continuation probabilities:

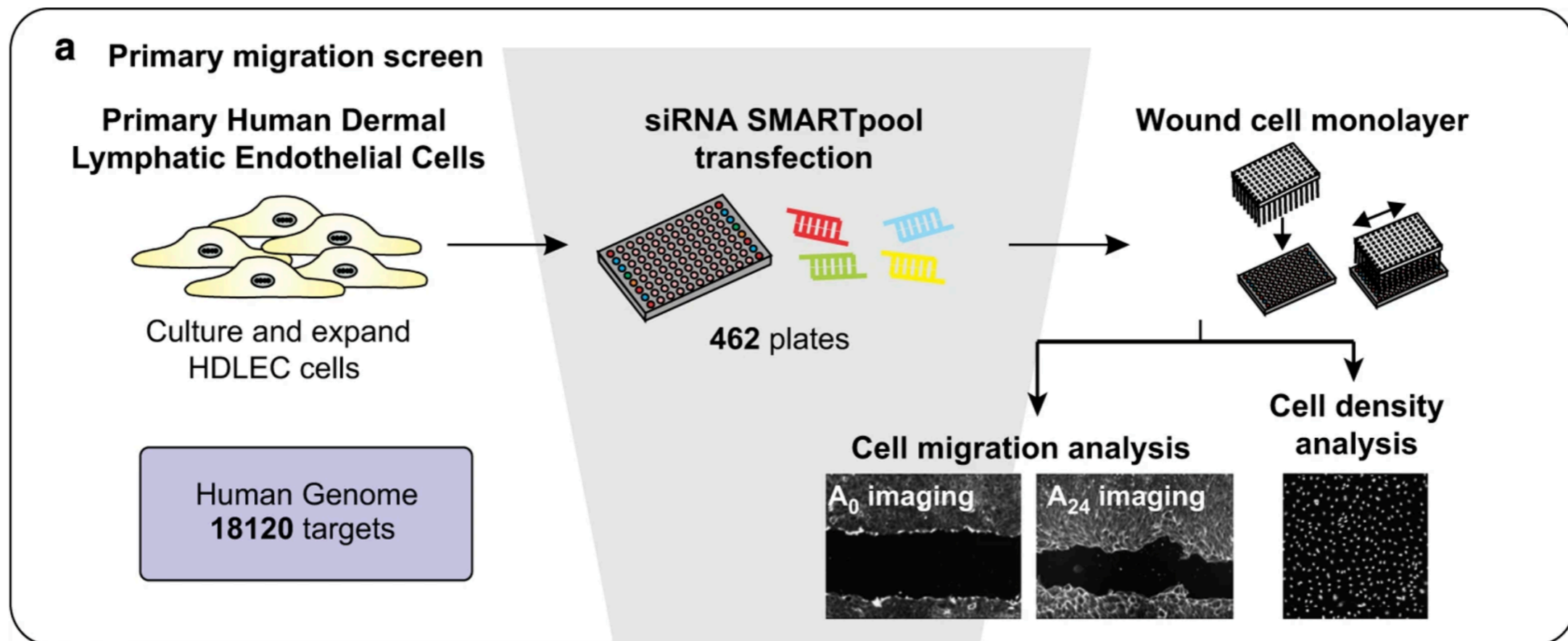


Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).

- Multifidelity ABC can provide time savings, through the combined use of high- and low-fidelity models.
- Can “learn” optimal continuation probabilities as the algorithm proceeds, separately controlling rates of checking early acceptance and early rejection.
- Rates of false positives and negatives can be reduced by generating the high-fidelity model output conditional on the low-fidelity model output.
- Enables smaller continuation probabilities and hence simulation cost.

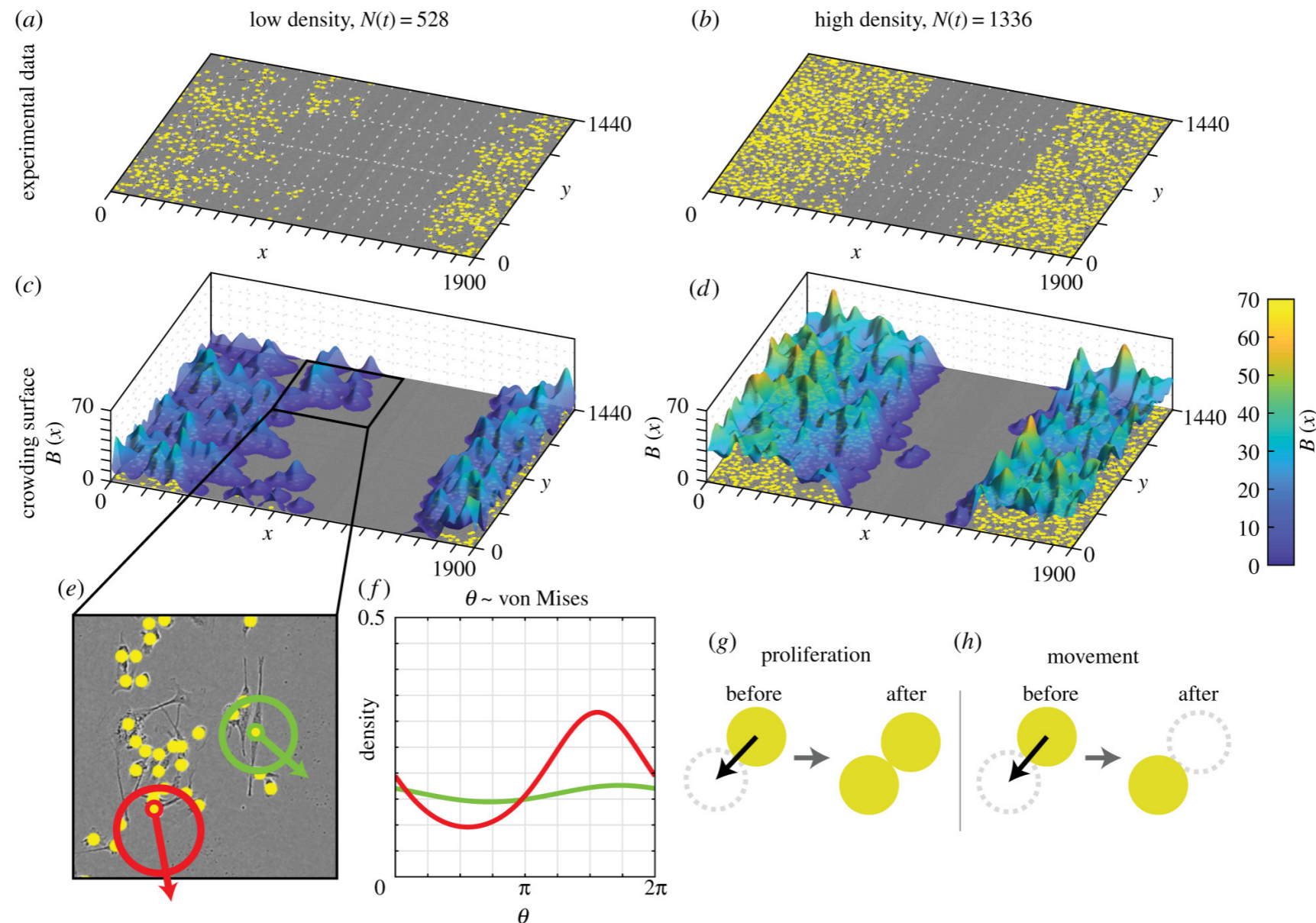
Prescott and Baker, *SIAM / ASA J. UQ* (2020) and (2021).

- Designed for use with high-throughput data i.e. large quantities of data.
- Motivated by genome-wide RNAi screen of endothelial cells.



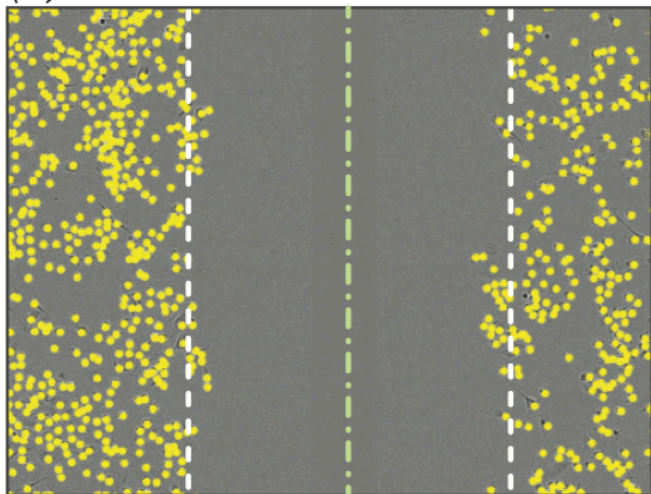
Williams *et al.* *Sci. Data* 4 (2017).

- Stochastic, off-lattice individual-based model of cell motility and proliferation - including density-dependent effects.

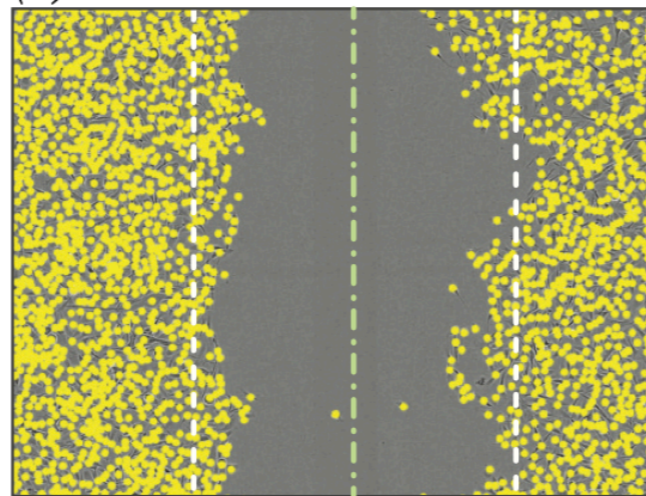


Browning *et al.* *J. Roy. Soc. Interface* 17 (2020).

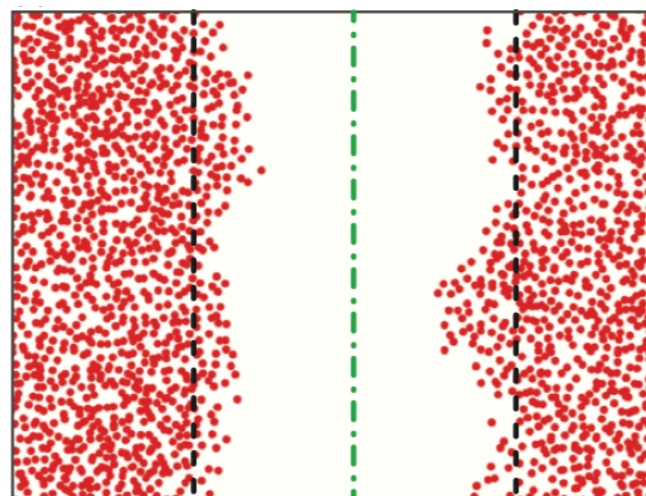
Data at t=0 hrs



Data at t=24 hrs



Model at t=24 hrs



Parameter inference using a Bayesian framework:

$$\mathbb{P}(\theta | \mathcal{D}) \propto \mathcal{L}(\mathcal{D} | \theta) \mathbb{P}(\theta)$$

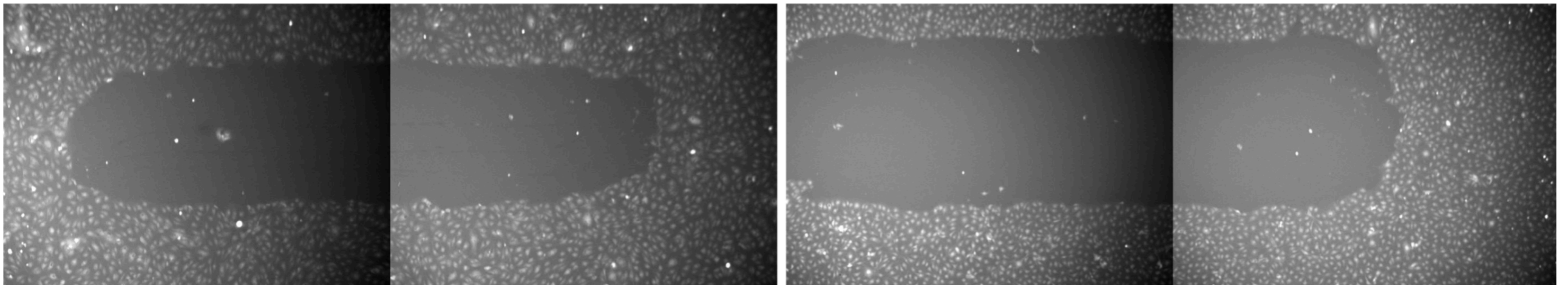
with

$$\theta = (m, p, d, \gamma_b, \gamma_m, \gamma_p)$$

Browning *et al.* *J. Roy. Soc. Interface* 17 (2020).

# What's the additional challenge?

- Large numbers of replicates ( $\sim 100$ ) for some knockdowns.
- Huge variability in the initial wound size / shape (initial condition).

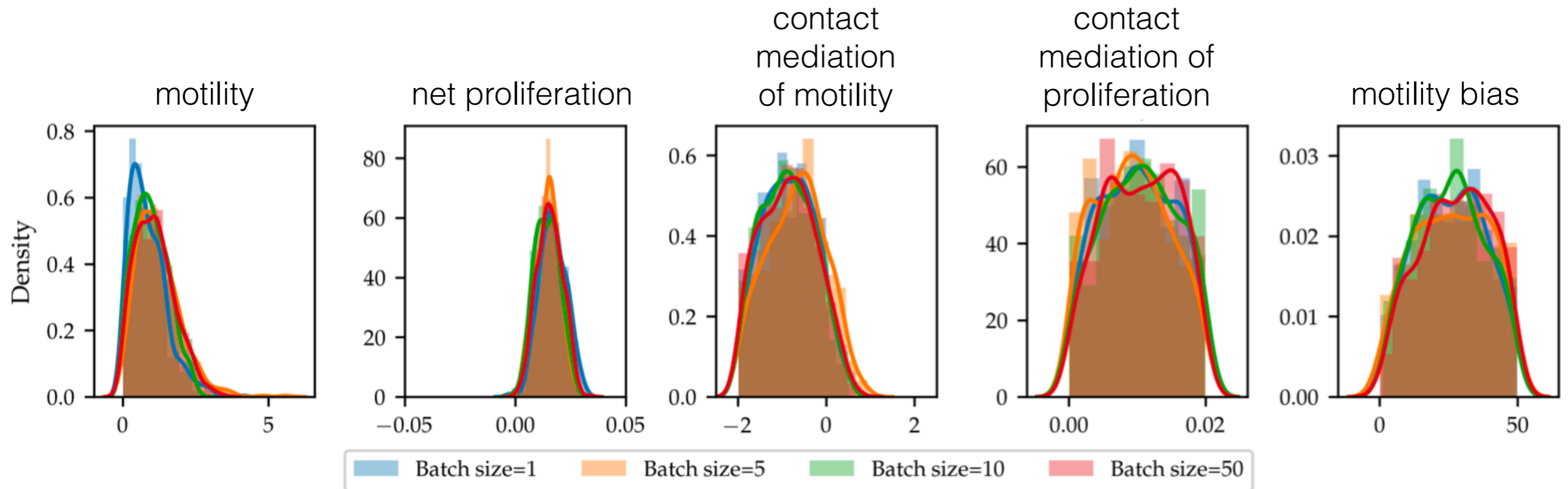


- Need to generate individual simulations / make individual comparisons for each replicate.

- A very large number of times:
  - sample parameter from prior distribution;
  - sample a **minibatch** of the data;
  - simulate model using this parameter - **do this for each sample from the minibatch individually**;
  - evaluate how close model output is to data using summary statistics and distance function - **do this for each sample from the minibatch individually**;
  - assign a weight to the parameter - depends on this distance.

Martina Perez, Sailem and Baker, *PLoS Comp. Biol.* (2022).

# Minibatch ABC - test results

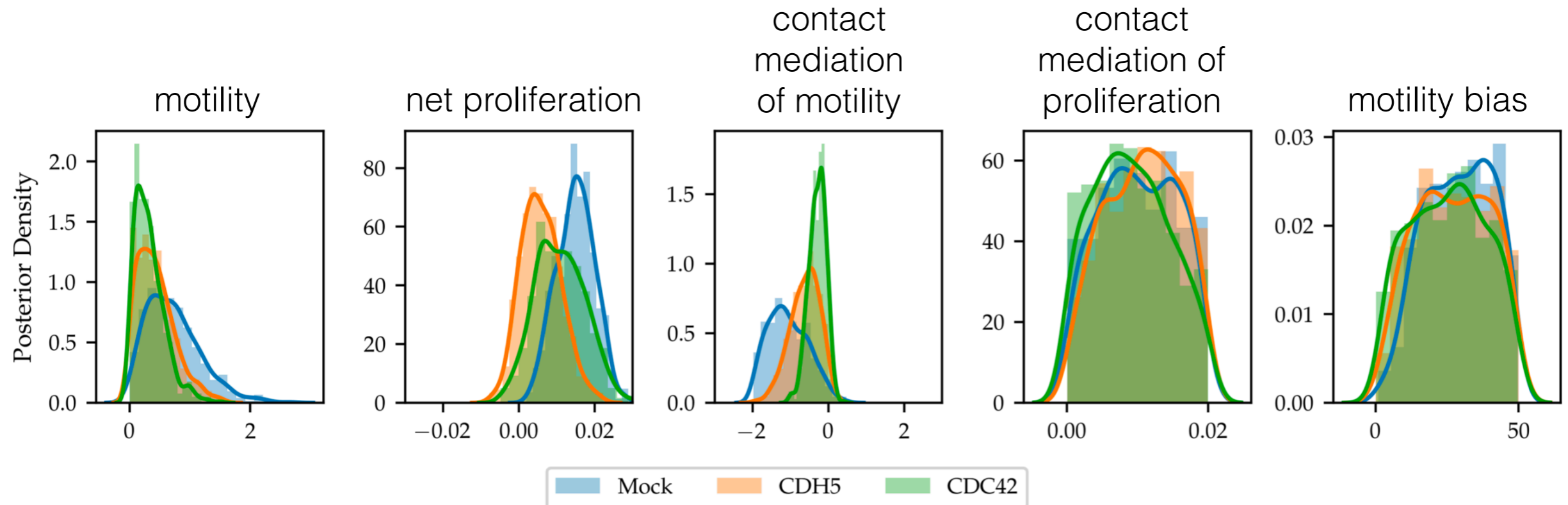


- Posterior distribution very similar over the different batch sizes.
- Confidence in estimates of motility, net proliferation, contact-mediation of motility parameters.

Martina Perez, Sailem and Baker, *PLoS Comp. Biol.* (2022).



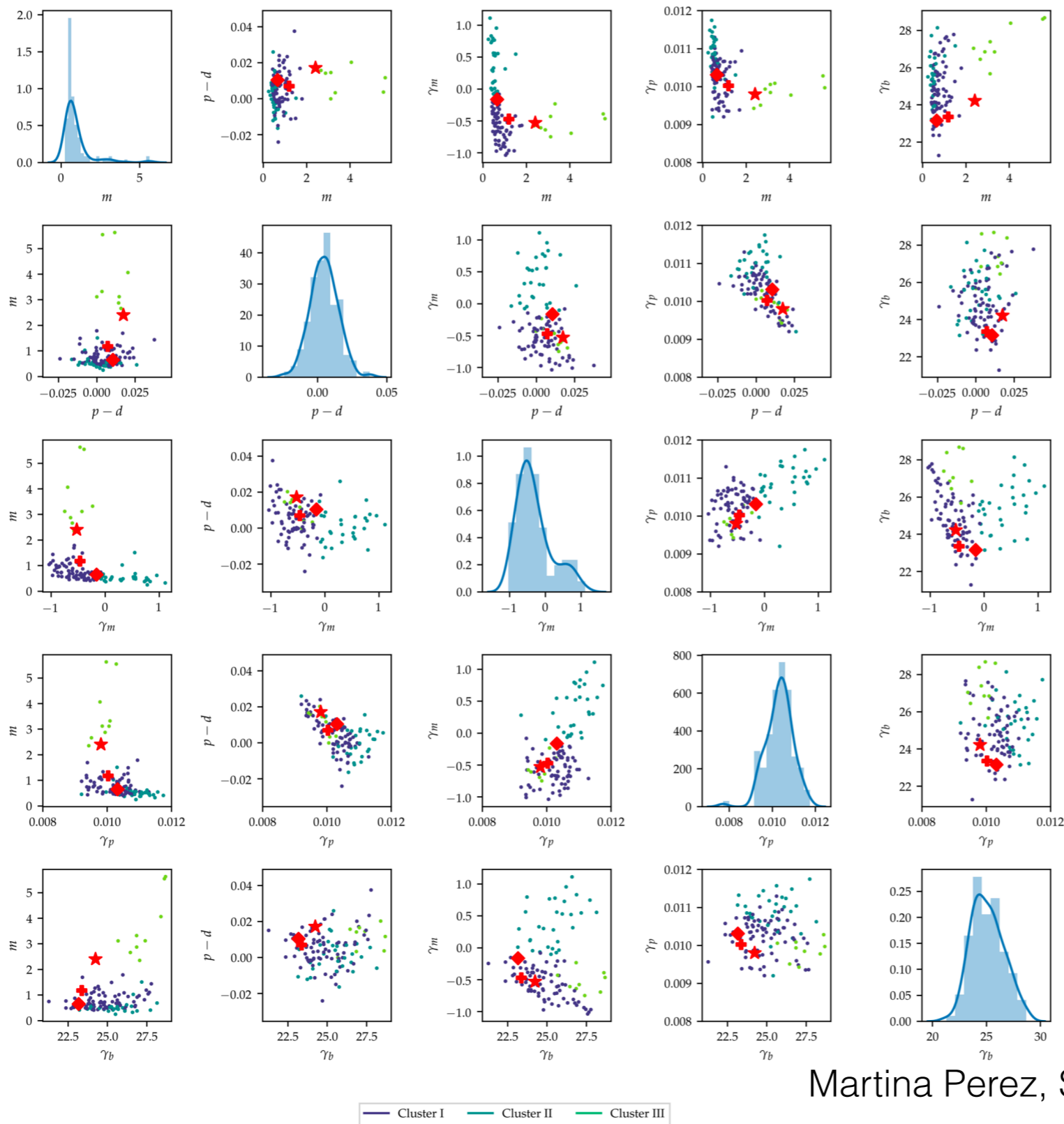
- Marginal posterior distributions



- Wildtype - motility strongly up-regulated in regions of high density compared to CDC42 and CDH5.
- Consistent with current understanding of the roles of CDC42 and CDH5.

Martina Perez, Sailem and Baker, *PLoS Comp. Biol.* (2022).

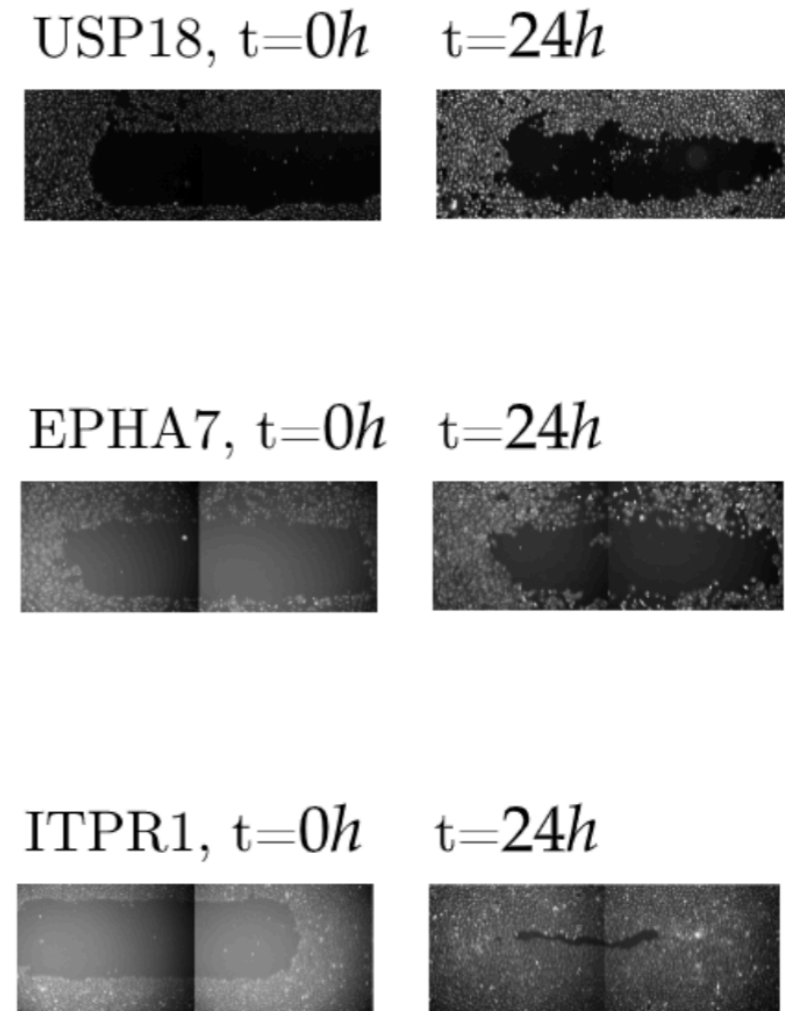
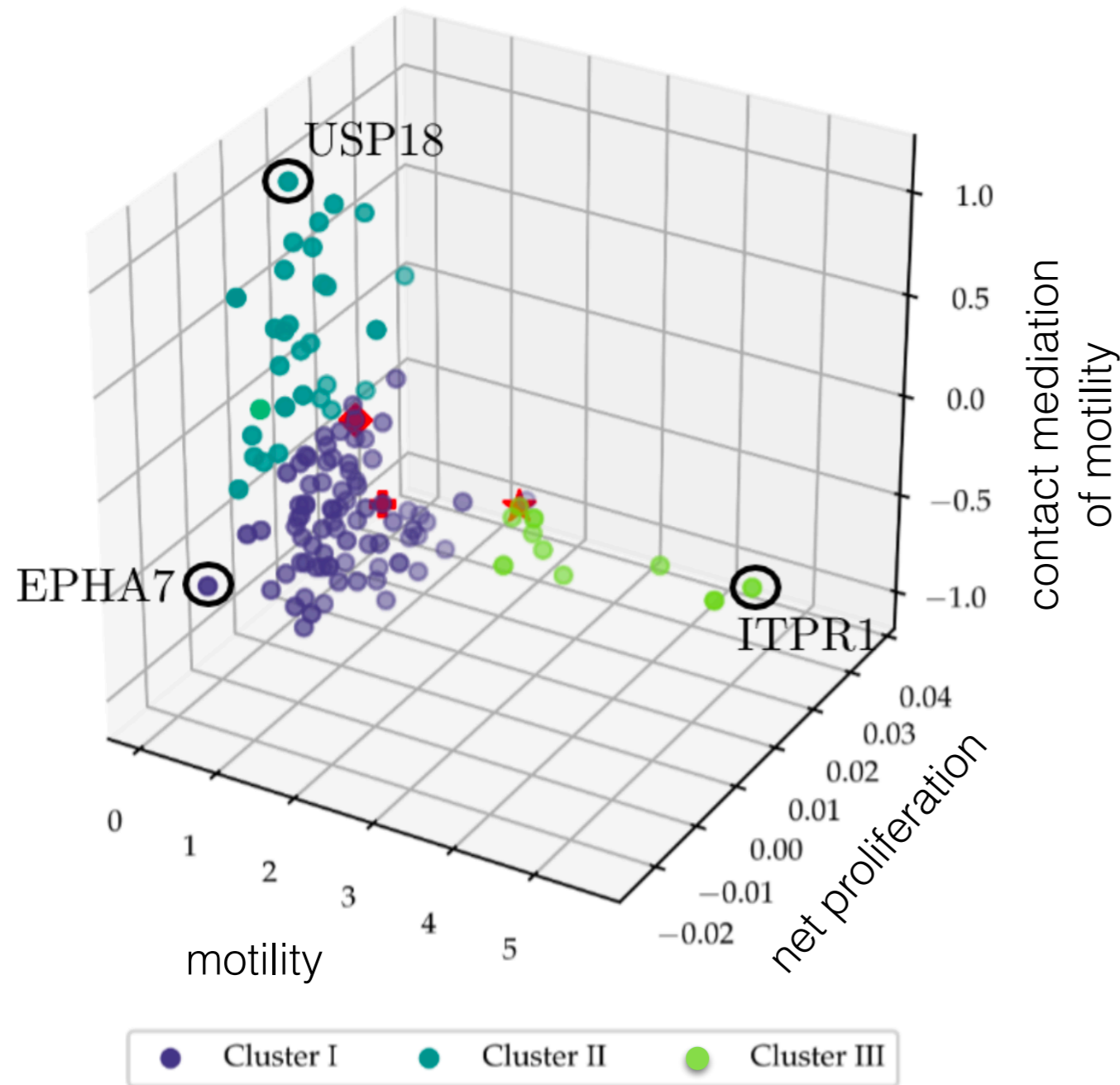
# Interpreting results of RNAi screen



- Repeat for the other genes in the RNAi screen...
- Plots of the posterior means for each knockdown.
- Used K-means clustering to establish three main clusters in phenotype space.

Martina Perez, Sailem and Baker, *PLoS Comp. Biol.* (2022).

# Interpreting results of RNAi screen



Martina Perez, Sailem and Baker, *PLoS Comp. Biol.* (2022).

- Possible to calibrate more complicated models to data, and use them to infer greater detail on the mechanisms driving wound closure in a range of gene knockdowns.
- Complex relationships between cell motility and proliferation drive wound closure.
- Mini-batch ABC provides a means to apply ABC approaches to high-throughput datasets, without huge computational costs.

- ABC is a fantastic tool for calibrating models to data since it relies only on forward simulation of the model.
- However, for modern mathematical biology studies the computational costs of naive forms of the method are prohibitive.
- Proposed three novel approaches to ABC - pre-conditioned, multi-fidelity and mini-batch.
- Importantly, ALL of these approaches can be combined with existing approaches e.g. ABC-SMC to provide further improvements.
- There's still much to do to optimise each approach!

# Acknowledgements



## Oxford

Tom Prescott  
Simon Martina-Perez

## Further afield

David Warne (QUT)  
Mat Simpson (QUT)



- First, need to integrate importance sampling into the multifidelity ABC framework:

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**Algorithm 5** Multifidelity ABC importance sampling (MF-ABC-IS)

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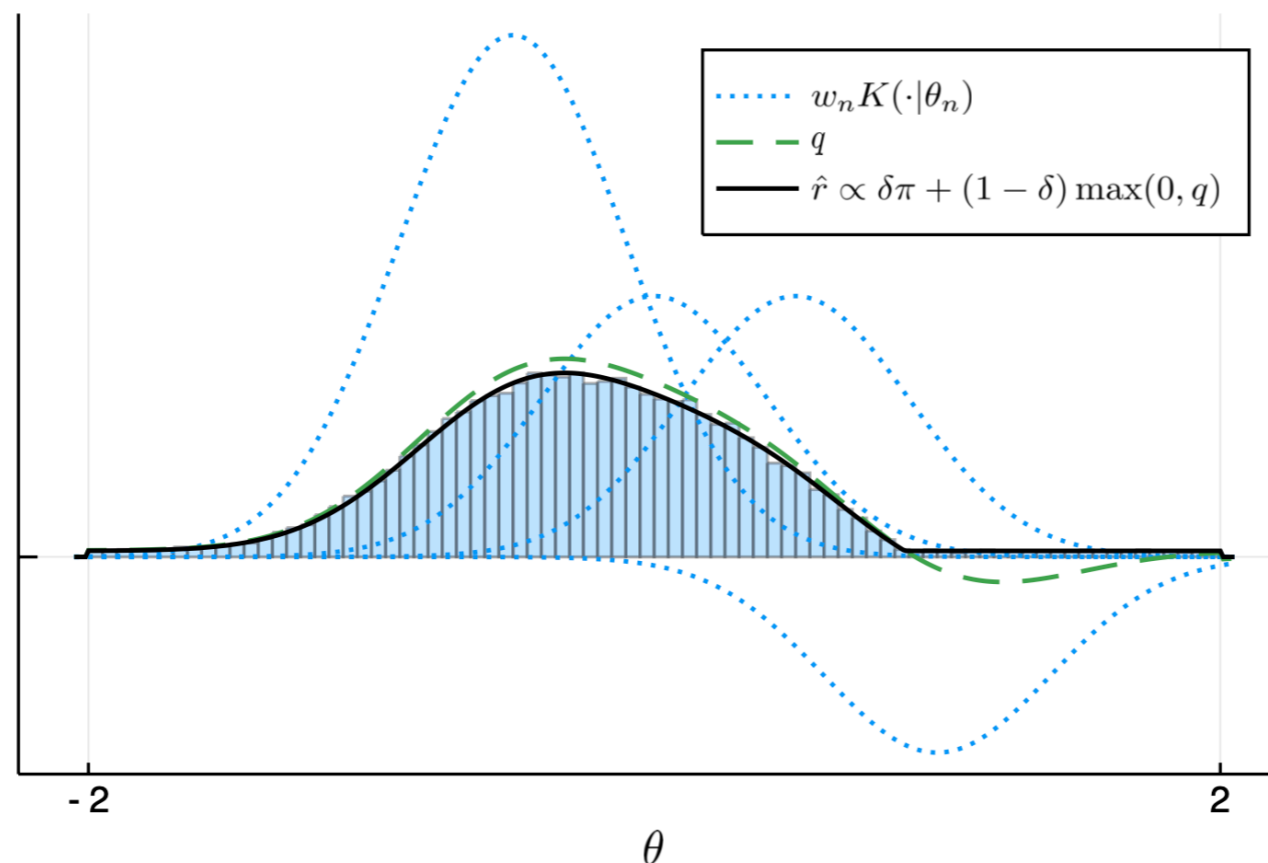
**Input:** Data  $y_{\text{obs}}$  and neighbourhood  $\Omega_\epsilon$ ; prior  $\pi$ ; models  $\tilde{f}(\cdot | \theta)$ ,  $f(\cdot | \tilde{y}, \theta)$ ; continuation probability function  $\alpha = \alpha(\tilde{y}, \theta)$ ; sample index  $n = 0$ ; importance distribution  $\hat{q}$  proposed by  $\theta_n$ .

**Output:**

- 1: repeat
- 2:   In
- 3:   Ge
- 4:   Si
- 5:   Se
- 6:   Generate  $u_n \sim \text{Uniform}(0, 1)$ .
- 7:   **if**  $u_n < \alpha(\tilde{y}_n, \theta_n)$  **then**
- 8:     Simulate  $y_n \sim f(\cdot | \tilde{y}_n, \theta_n)$ .
- 9:     Update  $w_n \leftarrow w_n + [\mathbb{I}(y_n \in \Omega_\epsilon) - w_n] / \alpha(\tilde{y}_n, \theta_n)$ .
- 10:   **end if**
- 11:   Update  $w_n \leftarrow [\pi(\theta_n) / q(\theta_n)] w_n$ .
- 12: **until**  $S = \text{true}$ .

**For SMC: how do we sample from the importance distribution, given the weights that result from multifidelity ABC can be negative?**

- Use defensive importance sampling, first defining a new (non-negative) importance distribution.



- Estimate continuation probabilities for each generation “on the fly”, using information from the previous generations.



- Kuramoto oscillator network:

$$\dot{\phi}_i = \omega_i + \frac{K}{M} \sum_{j=1}^M \sin(\phi_j - \phi_i)$$

angular velocities drawn  
from Cauchy distribution  
median -  $\omega_0$   
dispersion -  $\gamma$

- Low-fidelity model - based on tracking Daido order parameters:

$$Z_n(t) = \frac{1}{M} \sum_{j=1}^M \exp(in\phi_j)$$

assume

$$Z_n(t) = Z_1(t)^n$$

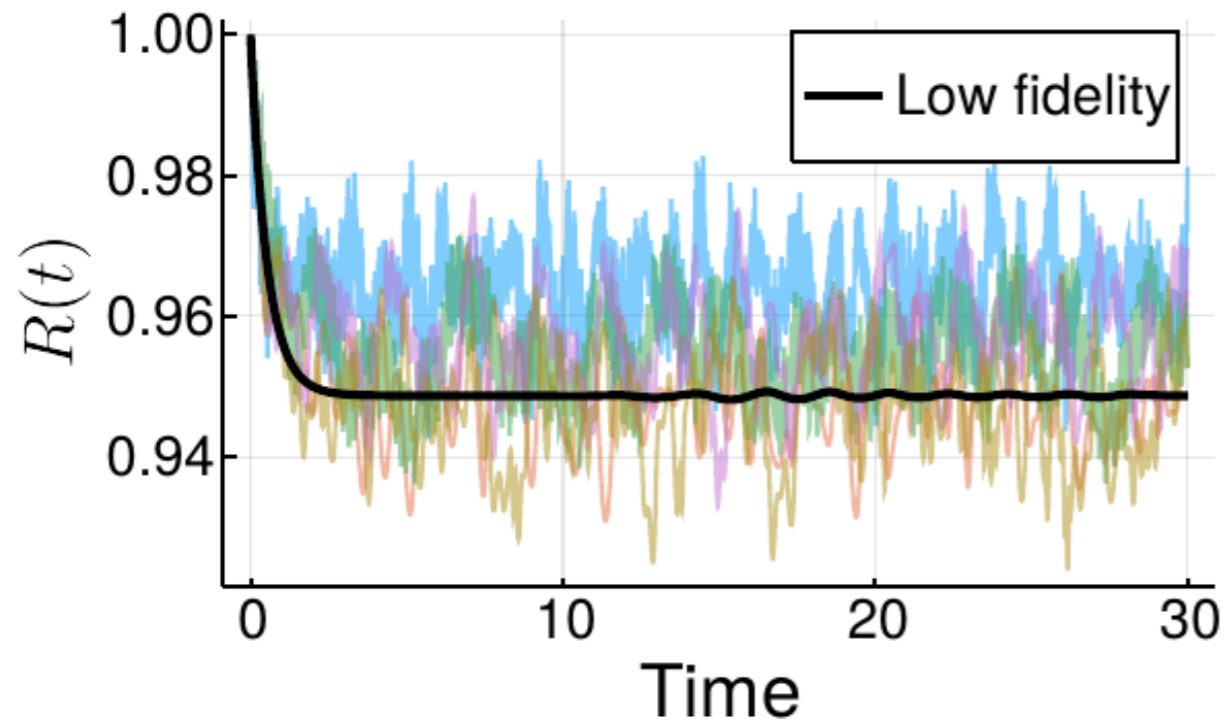
to get

$$\dot{\tilde{R}} = \left( \frac{K}{2} - \gamma \right) \tilde{R} - \frac{K}{2} \tilde{R}^3 \quad \text{(magnitude)}$$

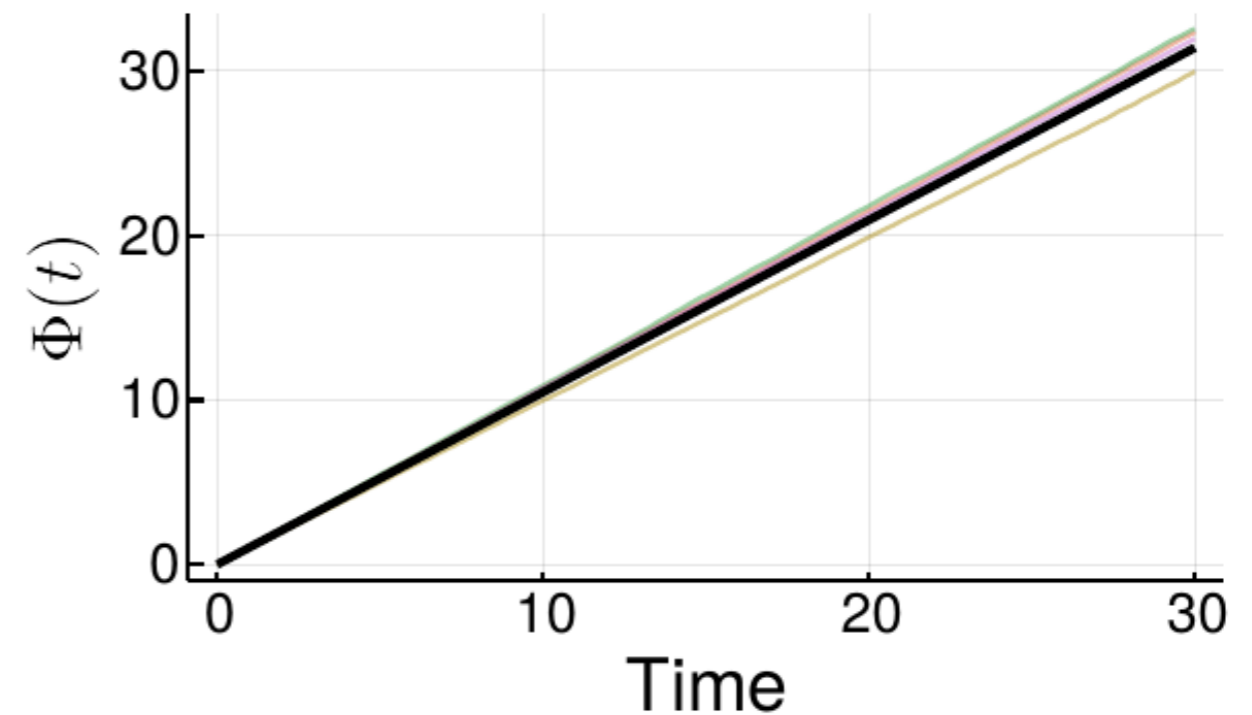
$$\dot{\tilde{\Phi}} = \omega_0 \quad \text{(phase)}$$

- Typical simulation output:

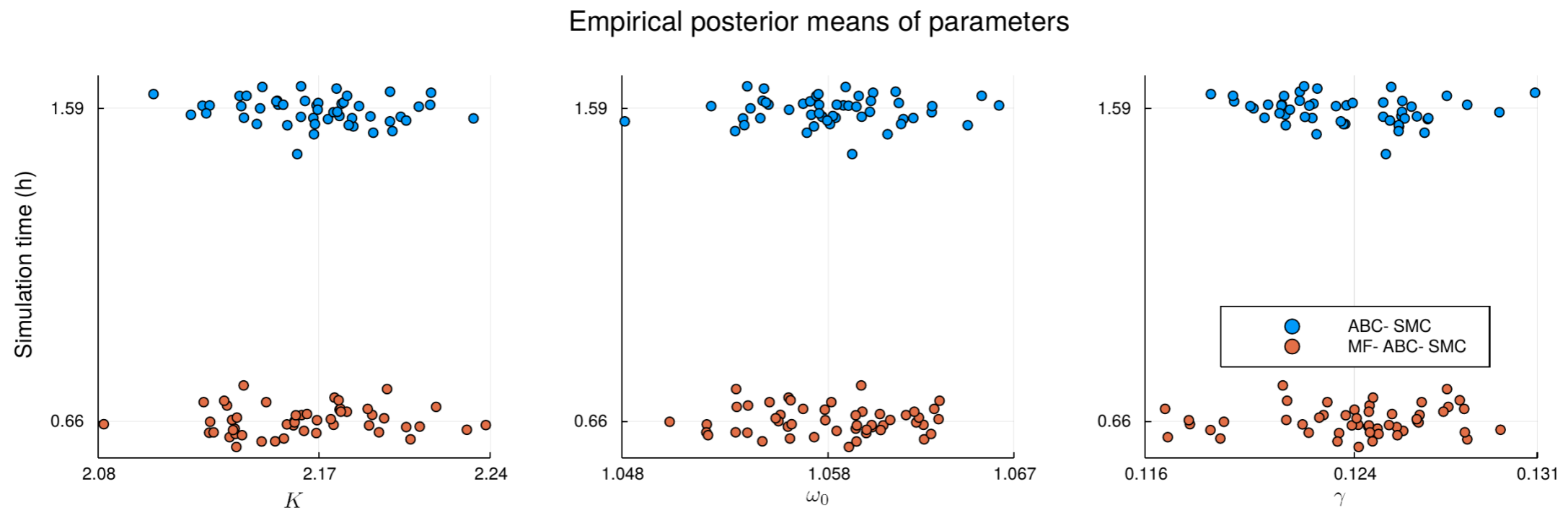
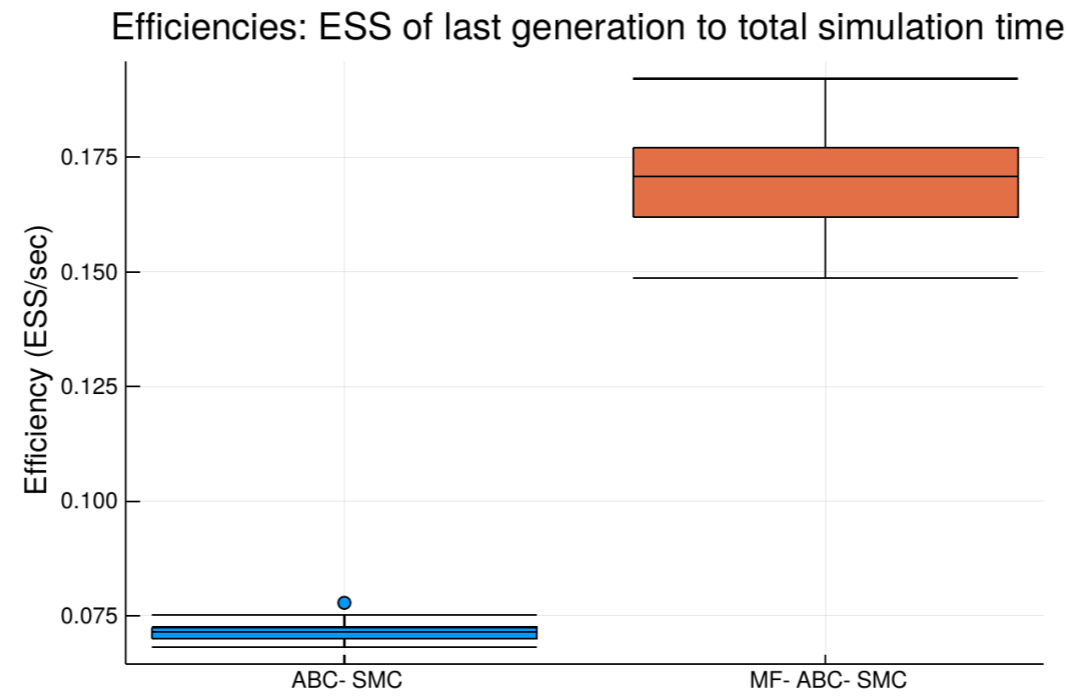
Kuramoto parameter: magnitude



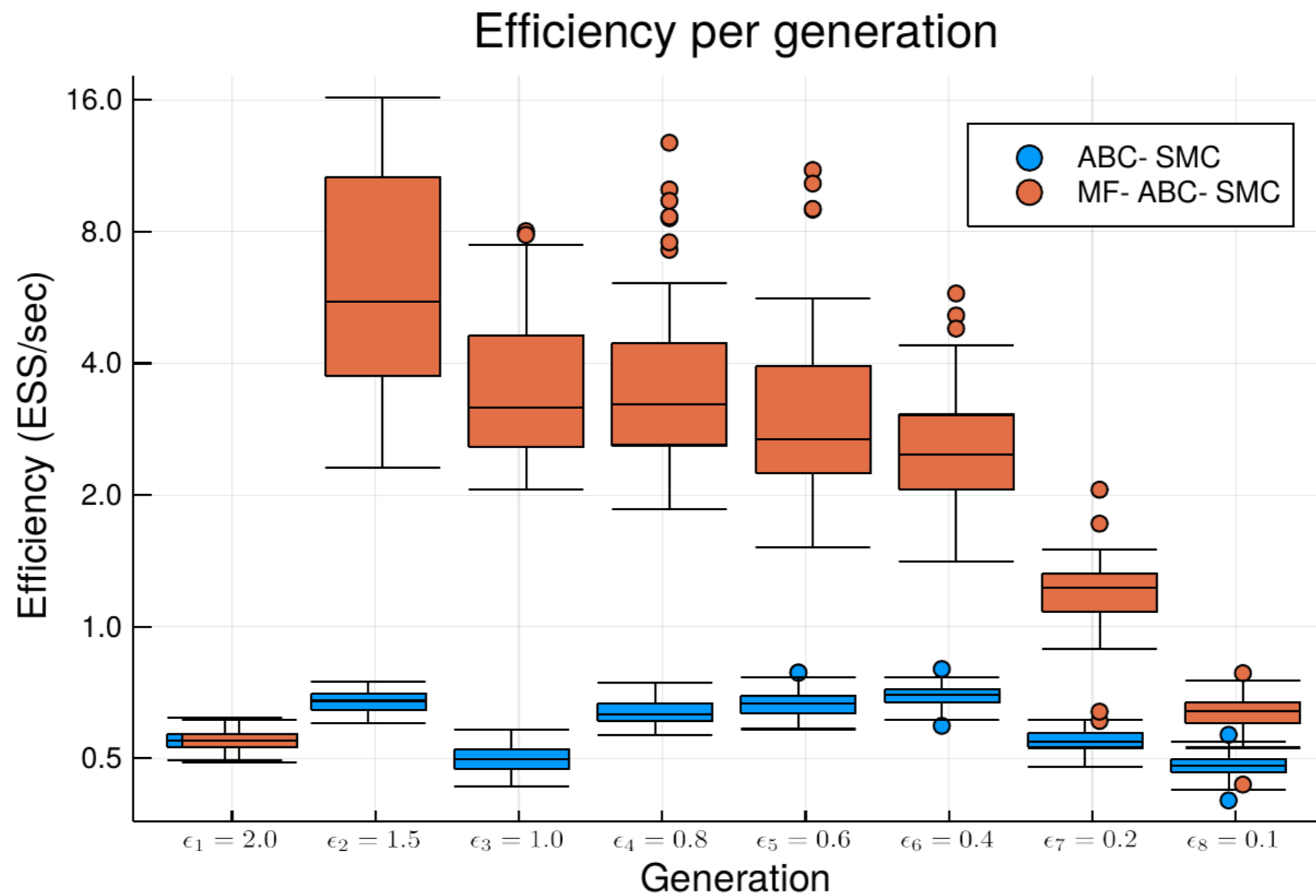
Kuramoto parameter: phase



# MF ABC SMC in action

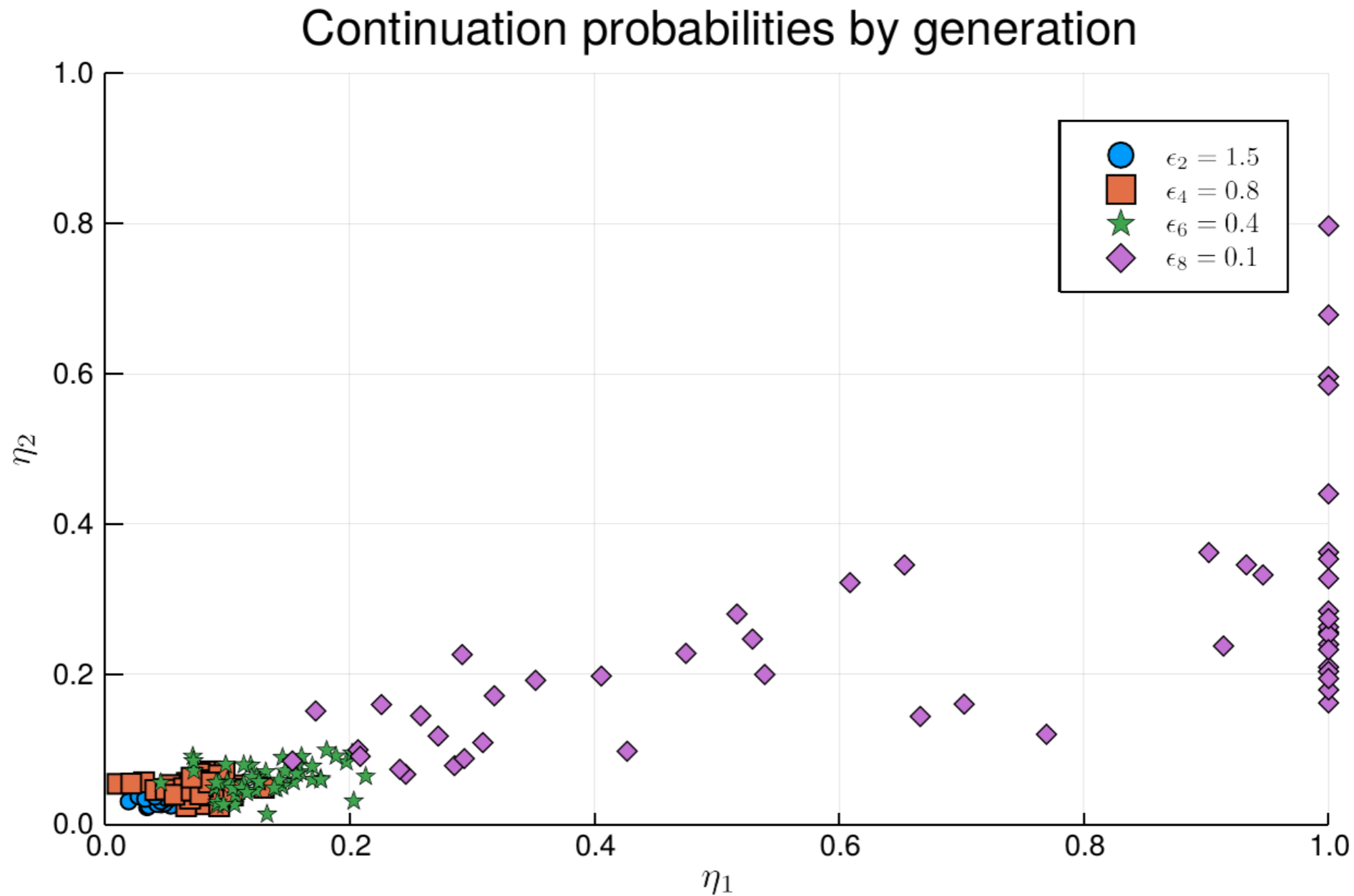


- Stopping criterion at each generation:  $ESS \geq 400$ .



# Towards multifidelity SMC-ABC

probability of requiring high-fidelity model  
simulation given low-fidelity model **far**



probability of requiring high-fidelity model  
simulation given low-fidelity model **close**